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## MOULDS AND MYCOTOXINS IN STORED MAIZE GRAINS

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**Abstract:** In this study the presence of moulds and mycotoxins in samples of stored maize grains in the period from October 2011 to September 2012 was investigated. Mycological analyses of whole and broken grains showed the presence of species from the genera *Alternaria*, *Aspergillus*, *Fusarium*, *Penicillium*, *Rhizopus* and others. Among the *Aspergillus* and *Fusarium* genera as potentially toxigenic fungi, *Aspergillus flavus* was identified with the highest percentage on broken grains (20.38%) whereas *F. verticillioides* was the predominant species in the whole maize grains (34.04%). In addition, it was obtained that tested samples of stored maize grains were 100% positive with aflatoxin B<sub>1</sub> (AFB<sub>1</sub>), zearalenone (ZON), deoxynivalenol (DON) and fumonisin B<sub>1</sub> (FB<sub>1</sub>) with an average concentration of 1.39 µg kg<sup>-1</sup>, 71.79 µg kg<sup>-1</sup>, 128.17 µg kg<sup>-1</sup>, and 1610.83 µg kg<sup>-1</sup>, respectively. A significant positive correlation was found between the moisture content and the presence of *Fusarium* spp. on the broken grains ( $r = 0.44$ ) and between the moisture content and the concentration of DON ( $r = 0.61$ ). However, a significant negative correlation was found between moisture content and FB<sub>1</sub> ( $r = -0.34$ ), and between the concentration of ZON and DON mycotoxins ( $r = -0.58$ ).

**Key words:** moulds, mycotoxins, storage, maize grains

## Introduction

Maize (*Zea mays* L.) is one of the most important sources of food for human and animal nutrition and raw materials for industrial processing. In our country, maize has been grown on about 1.2 million hectares with an average yield of 5.4 t ha<sup>-1</sup> and with a production of 6.5 million tons in 2011 (*Statistical Yearbook of Serbia, 2012*).

The nutritional value of stored maize grains could vary significantly due to the interaction between the physical, chemical and biological factors. The contamination of maize with fungi (moulds) and mycotoxins represents a major problem for its use in human and animal nutrition. Infection of grains in the field by fungi could result in the production of mycotoxins during cultivation, harvesting, storage, transport and processing. The most important species of fungi and mycotoxins that could contaminate maize grains are *Aspergillus flavus* and aflatoxins, *Fusarium verticillioides*, *F. proliferatum* and fumonisins and *F. graminearum* and trichothecenes and zearalenone (Chulze, 2010). Aflatoxin causes serious problem in many foods, but it is most abundant in maize and maize products, because maize could be infected even in the field under specific environmental conditions. Contamination of maize depends on the co-existence of susceptibility of hybrids and environmental conditions favourable for proliferation of mycotoxigenic fungi (Blandino et al., 2009).

Unfortunately, there are no direct measures for prevention of infection of maize grains with ear rot fungi. However, unfavourable conditions for the development of fungi and toxinogenesis could be provided by implementation of appropriate agricultural practices as preventive measures in the field. In addition, early spring planting of maize that extends the vegetation season and selection of hybrids with higher early vigor, tolerance to biotic and abiotic stresses could be considered as an important measures for reduction of pathogenic and toxigenic fungi, resulting in an increase of maize production (Lee et al., 2002; Blandino et al., 2009).

The most common genera of fungi identified in stored maize grains are *Aspergillus*, *Penicillium* and *Fusarium*. The proliferation of these fungi are stimulated with higher grains moisture content, higher temperature during storage, long storage period, intensive infection by fungi before storage and by higher activity of insects and mites. Therefore, it is important to identify the species of fungi in stored maize grains with special emphasis on mycotoxigenic species, which pose a potential risk to human and animal health (Castellari et al., 2010).

Therefore, the aim of this study was to identify the most important species of fungi with special focus on *Aspergillus* and *Fusarium* species as well as to quantify the associated mycotoxins in stored maize grains, intended for feeding of domestic animals (pigs, sheep and poultry).

## Materials and Methods

For mycological and mycotoxicological analysis of maize grains in the warehouse of the Institute for Animal Husbandry, Belgrade, samples were randomly taken according to the Commission Regulation (EC) No 401/2006 (European Commission, 2006) on every month during 12 months, from October

2011 to September 2012. Moisture content of maize grains was determined using a moisture analyzer (OHAUS MB35, USA).

Twelve different samples of maize were tested by mycological methods, where in each sample 200 whole and 200 broken grains were analyzed. The grains were first rinsed by tap water, then disinfected in a 1% solution of sodium hypochlorite (NaOCl) 3-5 minutes and distributed on the surface of water agar (WA) in Petri dishes (5 grains per Petri dishes) and incubated in a thermostat for 5-7 days at 25 °C. Identification of colonies of fungi that overgrew the maize grains was done by microscopic examination of mycelium and spores, according to *Burgess et al. (1994)* and *Watanabe (1994)*. The frequency of individual species was calculated per sample according to the following equation: (Number of grains in which the fungus was detected / Total number of grains) x 100.

Detection of aflatoxin B<sub>1</sub> (AFB<sub>1</sub>), zearalenone (ZON), deoxynivalenol (DON) and fumonisin B<sub>1</sub> (FB<sub>1</sub>) in a total of 12 samples was performed according to enzyme-linked immunosorbent assay (ELISA). Each sample was divided into two sub-samples, one with whole, and the other with broken grains. Individual sub-sample was milled in an analytical mill (IKA A11, Germany) and stored in a refrigerator at 4 °C, prior to the analyses. Five grams of subsample was mixed with 1 g of NaCl and homogenized in 25 ml of 70% methanol in a 250 ml Erlenmeyer flask on the orbital shaker (GFL 3015, Germany) for 30 minutes. Homogenate was filtered through a Whatman filter paper 1. The filtrate was further analyzed according to the manufacturer's instructions Celery techno ® ELISA kits. Absorbance was measured at a wavelength of 450 nm on an ELISA reader spectrophotometer (Biotek EL x 800TM, USA).

**Statistical analyses.** The correlation between the individual values obtained for the moisture content, the frequency of moulds and mycotoxins concentration was performed using Pearson's correlation coefficient.

## Results

The moisture content of the samples of stored maize was in the range of 9.26 to 12.58% with an average moisture content of 11.02%.

In the stored maize grains it was identified several fungal genera, as followed: *Alternaria*, *Aspergillus*, *Fusarium*, *Penicillium*, *Rhizopus* and others. By observing the average presence of the isolated fungi in the both grains (whole and broken) it was obtained that most of the genera were more frequent in the broken compared to whole grains, except *Fusarium*, whose representatives were more frequent in the whole grains (38.6%) compared to the broken grains (25.3%). Toxigenic species of the genus *Aspergillus* were present at levels ranging from 0 (March, April, and June 2012) to 39% (August 2012) in the whole maize grains, with an average frequency of 14.5% for all investigated months. These species were present in the range from 0 (April 2012) to 68.5% (July 2012) in the broken

maize grains with an average of 23.7% for all investigated months. *Fusarium* species were present in the range from 3 (October 2011) to 97.5% (June 2012) in the whole grains and 2.5 (January 2012) to 77% (June 2012) on a broken maize grains (Table 1).

Weak positive correlation was found between moisture content and frequency of *Aspergillus* spp. ( $r = 0.17$ ) and *Fusarium* spp. ( $r = 0.18$ ) in the whole maize grains. Statistically significant correlations were found between the moisture content and *Fusarium* spp. ( $r = 0.44$ ) to a broken maize grains and statistically insignificant negative correlation ( $r = -0.09$ ) between the moisture content and *Aspergillus* spp.

**Table 1. Frequency of fungal genera in stored maize grain samples during investigated period**

Sample <sup>a</sup>	Fungal genera											
	<i>Alternaria</i>		<i>Aspergillus</i>		<i>Fusarium</i>		<i>Penicillium</i>		<i>Rhizopus</i>		Other	
	W	B	W	B	W	B	W	B	W	B	W	B
	Percentage of infected grains											
1	2.5	15.5	4.5	12	3.0	36.5	4.5	35.0	47.5	31.5	14.0	0.0
2	0.0	1.5	8.0	19	12.0	22.5	0.0	18.0	83.0	73.0	0.0	0.0
3	4.5	0.0	11.0	18.5	52.5	45.0	2.5	9.5	22.0	55.0	4.0	0.0
4	0.0	7.5	23.0	50	36.5	2.5	1.5	24.5	37.5	63.0	1.5	1.0
5	14	2.5	14.0	20.5	20.5	27.5	14.0	48.5	45.5	29.5	1.0	0.0
6	32	43.0	0.0	20.5	45.5	14.0	0.0	21.5	22.5	37.5	13.5	10.5
7	0.0	23.5	0.0	0.0	35.0	17.0	39.5	46.0	11.0	18.5	1.0	0.0
8	0.0	0.0	5.5	3.5	46.0	11.5	34.5	67.5	23.0	47.0	0.0	0.0
9	0.0	0.0	0.0	5.5	97.5	77.0	3.0	20.0	3.5	20.0	0.0	5.5
10	2.5	0.5	37.0	68.5	9.5	4.0	13.5	5.0	2.0	27.0	0.5	0.0
11	0.0	5.0	39.0	25	37.0	29.0	1.5	6.0	30.0	5.5	0.5	12.5
12	0.5	0.0	32.0	41	40.5	16.5	1.5	2.0	3.5	44.5	21.5	16.0
Average	4.7	8.3	14.5	23.7	38.6	25.3	9.7	25.3	27.6	37.7	4.8	3.8

<sup>a</sup>The dates when the samples were collected: 1 - October 2011; 2 - November 2011; 3 - December 2011; 4 - January 2012; 5 - February 2012; 6 - March 2012; 7 - April 2012; 8 - May 2012; 9 - June 2012; 10 - July 2012; 11 - August 2012; 12 - September 2012;

W - Whole maize grains; B - Broken maize grains

Statistical high positive correlation was found between contamination of whole and broken grains with *Aspergillus* spp. ( $r = 0.79$ ), *Penicillium* spp. ( $r = 0.74$ ) and *Alternaria* spp. ( $r = 0.72$ ) whereas it was significant with *Fusarium* spp. ( $r = 0.61$ ) and *Rhizopus* spp. ( $r = 0.51$ ). Interrelationship of identified species of fungi in the whole and broken grains was generally negatively correlated, which was not statistically significant, except in the case of *Aspergillus* spp. and *Fusarium* spp. ( $r = -0.52$ ) and *Aspergillus* spp. and *Penicillium* spp. ( $r = -0.59$ ) on the broken grains and *Fusarium* spp. and *Rhizopus* spp. on the whole grains ( $r = -0.52$ ). The positive correlation that was not statistically significant, was found only between *Aspergillus* spp. and *Rhizopus* spp. ( $r = 0.16$ ), and *Alternaria* spp. and

*Penicillium* spp. ( $r = 0.14$ ) on a broken grains as well as between *Rhizopus* spp. and *Alternaria* spp. ( $r = 0.02$ ) on the whole grains.

From the tested grains, *A. flavus*, *A. parasiticus* and *A. niger* were isolated as the main species of the genus *Aspergillus*, whereas *F. graminearum*, *F. proliferatum*, *F. subglutinans* and *F. verticillioides* were isolated as species of the genus *Fusarium*. Generally, *Aspergillus* species were more presented on the broken grains, compared with whole grains of the stored maize (Table 2). From the *Aspergillus* species, *A. flavus* was the most presented on both the whole (12.3%), and the broken grains (20.38%). Of all identified species, including *Fusarium* spp., the most common species was *F. verticillioides*, on the whole (34.4%) and broken maize grains (23.9%).

**Table 2. Frequency of *Aspergillus* spp. and *Fusarium* spp. in investigated stored maize grain samples**

Fungal species	Percentage of infected grains	
	W	B
<i>Aspergillus flavus</i>	12.25	20.38
<i>Aspergillus parasiticus</i>	0.08	0.33
<i>Aspergillus niger</i>	2.17	2.96
<i>Fusarium graminearum</i>	0.17	0.33
<i>Fusarium proliferatum</i>	0.29	0.0
<i>Fusarium subglutinans</i>	4.13	1.0
<i>Fusarium verticillioides</i>	34.04	23.92

W – Whole maize grains; B – Broken maize grains

Concerning the presence of AFB<sub>1</sub>, ZON, DON and FB<sub>1</sub>, it was detected in all tested samples with an average concentrations of 1.39 µg kg<sup>-1</sup>, (range 0.33 to 2.40 µg kg<sup>-1</sup>), 71.79 µg kg<sup>-1</sup> (range 15.44-188.05 µg kg<sup>-1</sup>), 128.17 µg kg<sup>-1</sup> (range 41-226 µg kg<sup>-1</sup>) and 1610.83 µg kg<sup>-1</sup> (range 880-2950 µg kg<sup>-1</sup>), respectively (Table 3).

A significant positive correlation has only been found between moisture content and DON ( $r = 0.61$ ), while for the other mycotoxins tested this correlation was negative for AFB<sub>1</sub> ( $r = -0.07$ ), ZON ( $r = -0.25$ ) and FB<sub>1</sub> ( $r = -0.34$ ). The correlation between the concentrations of ZON and DON was significantly negative ( $r = -0.58$ ), and between AFB<sub>1</sub> and FB<sub>1</sub> was statistically insignificant negative ( $r = -0.08$ ).

**Table 3. Concentration of aflatoxin B<sub>1</sub> (AFB<sub>1</sub>), zearalenone (ZON), deoxynivalenol (DON) and fumonisin B<sub>1</sub> (FB<sub>1</sub>) in investigated stored maize grain samples**

Item	AFB <sub>1</sub>	ZON	DON	FB <sub>1</sub>
Sample size <sup>a</sup>	12/12	12/12	12/12	12/12
Incidence (%)	100	100	100	100
Range (µg kg <sup>-1</sup> )	0.33-2.40	15.44-188.05	41-226	880-2950
Mean <sup>b</sup> (µg kg <sup>-1</sup> )	1.39	71.79	128.17	1610.83

<sup>a</sup> Number of positive samples/Number of total samples

<sup>b</sup> Mean concentration in positive samples

## Discussion

In this study the presence of potentially toxigenic fungi of the genera *Aspergillus* (*A. flavus*) and *Fusarium* (*F. verticillioides*) on the stored maize grains was performed. The obtained results are in agreement with previous investigations reported by Hell (2003), Krnjaja et al. (2007) and Amadi and Adeniyi (2009). By mycological testing of 86 samples of stored maize, originating from different farms in two locations in Kenya, it was found that the most frequently isolated species were from the genera *Aspergillus* (35.8%), and *Fusarium* (15.5%), followed by *Penicillium* (9.2%), *Rhizopus* (5.3 %) and others (34.4%) (Bii et al., 2012).

The co-occurrence of toxigenic fungi is not uncommon feature. During the study period it was found the co-occurrence of identified genera *Aspergillus*, *Fusarium*, *Penicillium* and *Rhizopus*, especially on broken maize grains. Also, Remešova et al. (2007) isolated the same genera of fungi in healthy and damaged maize, but with different frequency. In the study of Miller (1993) it was also found frequent joint occurrence of *A. flavus* and *F. verticillioides* on maize. It has already been reported that damage of maize germ has the greatest impact on the proliferation of moulds (Tuite et al., 1985).

Although in the tested samples almost all fungi were identified, their ratio was still generally negatively correlated, both in the whole, and the broken maize grains. Contrary to wheat, where statistically highly significant negative correlation between *Fusarium* spp. and *Alternaria* spp. was found (Lević et al., 2012) on maize grains this negative correlation was not statistically significant.

Aflatoxins and fumonisins, synthesized mainly by *A. flavus* and *F. verticillioides*, respectively, are among the most important mycotoxins that can cause economic losses in maize production. In this research, the co-occurrence of aflatoxin B<sub>1</sub> with fumonisin B<sub>1</sub> (100%) was found, which is similar to the observations of Kimanya et al. (2008), Sun et al. (2011) and Krnjaja et al. (2013).

All of the samples in this study were positive on the presence of *Fusarium* toxins, fumonisin B<sub>1</sub>, deoxynivalenol and zearalenone which is in accordance with the results of Lazzari (1994). Because there are favourable conditions for the growth and development of these fungi and mycotoxin contamination of maize grains, in Serbia it is of outmost importance to implement the preventive measures to reduce the risk of these contaminants, especially in years when weather conditions are suitable for their development. The weather conditions before and during the maize harvest could affect the safety of grains during storage. In the period of maize silking, high ambient temperature and high humidity could favour infection of grains with *Aspergillus* species particularly by *Aspergillus flavus* and mycotoxins. *Fusarium* ear rot of maize caused mostly by *F. verticillioides* and *F. proliferatum* is also stimulated by warm weather, dry seasons and increased damage of ear by insects (Miller, 1995). Drought, inadequate nutrition of plants, other agents of plant diseases, insects, weeds and excessive plant populations can

cause stress in plants and facilitate the infection of maize grains by toxigenic fungi. Sowing the right (adapted) hybrids, optimum plant nutrition, irrigation and insect control would certainly help to reduce mycotoxins contamination. Early harvesting and artificial drying reduces the occurrence of mycotoxins contributed the prevention of grain breakage and protect grain from insect pests (Bruns, 2003). Good ventilation of warehouse has also been one of the most important preventive measures for the reduction of mycotoxins production (Jakic-Dimic et al., 2011).

Contamination of maize by mycotoxins has been increasing worldwide, as a result of climate change, growing of high-yielding hybrids susceptible to infection with toxigenic fungi and accumulation of mycotoxins in crop products, especially in wheat. Some mycotoxins could be synthesized in maize before harvest but their concentration may increase after harvest during the storage period and further in the food chain. Preventive measures, such as fast drying of maize for the medium and long-term storage in hygiene maintained warehouses, without the presence of insects and microorganisms, and proper regulation of grains moisture content, could significantly reduce the mycotoxins contamination of maize grains.

## Conclusion

Based on the obtained results it can be concluded that the potentially toxigenic species of fungi from the genera *Aspergillus* and *Fusarium* were significantly presented in the stored maize grains. During the storage period (October 2011 - September 2012) *Aspergillus* spp. was the most frequently presented in August (39%) on the whole grains and in July (68.5%) on the broken grains, while *Fusarium* spp. was most presented in June on the both, whole (97.5%) and broken maize grains (77%). *F. verticillioides* was the most common species with an average frequency of 23.92% on broken and 34.04% on the whole maize grains. In addition, *A. flavus* was generally less presented on the whole (12.25%) than on the broken grains (20.38%).

Generally, it can be concluded that despite the significant presence of toxigenic species on the maize grains, the concentration of mycotoxins AFB<sub>1</sub>, ZON, DON and FB<sub>1</sub> has not exceed the maximum allowed concentrations prescribed by Regulation on the quality of animal feed (Article 99) (*Official Gazette of the Republic of Serbia*, 2010). These results indicated that the stored maize was suitable for feeding of farm animals (pigs, sheep and poultry).

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## Plesni i mikotoksini u uskladištenom kukuruзу

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### Rezime

U radu je ispitivano prisustvo plesni i mikotoksina u uzorcima zrna uskladištenog kukuruза u periodu od oktobra 2011. do septembra 2012. godine. Mikološkim analizama celog i slomljenog zrna kukuruза ustanovljeno je prisustvo vrsta iz rodova *Alternaria*, *Aspergillus*, *Fusarium*, *Penicillium*, *Rhizopus* i drugih. Od potencijalno toksigenih vrsta iz rodova *Aspergillus* i *Fusarium*, identifikovane su u najvećem procentu *A. flavus* na slomljenom (20,38%) i *F. verticillioides* na celom zrnju kukuruза (34,04%). Ispitivani uzorci uskladištenog kukuruза bili su 100% pozitivni sa aflatoksinom B<sub>1</sub> (AFB<sub>1</sub>), zearalenonom (ZON), deoksivalenolom (DON) i fumonizinom B<sub>1</sub> (FB<sub>1</sub>) sa prosečnim koncentracijama 1,39 µg kg<sup>-1</sup>, 71,79 µg kg<sup>-1</sup>, 128,17 µg kg<sup>-1</sup> i 1610,83 µg kg<sup>-1</sup>, respektivno.

Statistički značajna pozitivna korelacija ustanovljena je između sadržaja vlage i prisustva *Fusarium* spp. na slomljenom zrnju kukuruза ( $r = 0,44$ ), kao i između sadržaja vlage i koncentracije DON ( $r = 0,61$ ). Statistički značajna negativna korelacija ustanovljena je između sadržaja vlage i FB<sub>1</sub> ( $r = -0,34$ ), kao i između koncentracija ZON i DON mikotoksina ( $r = -0,58$ ).

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